

Cassane- and Norcassane-Type Diterpenes from *Caesalpinia crista* of Indonesia and Their Antimalarial Activity against the Growth of *Plasmodium falciparum*

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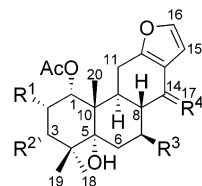
The CH₂Cl₂ extract of the seed kernels of *Caesalpinia crista*, which exhibited promising antimalarial activity against *Plasmodium berghei*-infected mice in vivo, was examined and resulted in the isolation of seven new furanocassane-type diterpenes [caesalpinins C–G (**1–5**) and norcaesalpinins D and E (**6, 7**)] together with norcaesalpinins A–C (**8–10**) and 11 known compounds (norcaesalpinins A–C, 2-acetoxy-3-deacetoxycaesaldekarin e, caesalmin B, caesaldekarin e, caesalpin F, 14(17)-dehydrocaesalpin F, 2-acetoxycaesaldekarin e, 7-acetoxybonducellpin C, and caesalmin G). Their structures were determined on the basis of spectroscopic analysis. The isolated diterpenes showed significant dose-dependent inhibitory effects on *Plasmodium falciparum* FCR-3/A2 growth in vitro. Their IC₅₀ values ranged from 90 nM to 6.5 μM, and norcaesalpinin E (**7**) showed the most potent inhibitory activity (IC₅₀, 90 nM).

Malaria is a parasitic disease affecting 200–300 million people in the tropical and subtropical regions of the world and claims the lives of approximately three million people each year.¹ The appearance of drug-resistant *Plasmodium falciparum* since 1960 has made the treatment of malaria increasingly problematic, and apparently the battle against malaria has not been successful. From the historical discovery of quinine from the *Cinchona* tree and the recent discovery of artemisinin from *Artemisia annua* L. (Asteraceae), there is anticipation that new leads may emerge from other tropical plant sources.

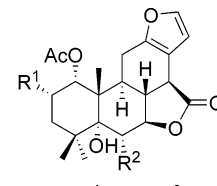
Caesalpinia crista Linn. (Fabaceae) is a popular traditional medicinal plant and is widely distributed throughout the tropical and subtropical regions of Southeast Asia. In Indonesia, it is commonly known as “Bagore”, and a decoction of the roots has been used as a tonic and for the treatment of rheumatism and backache.² Its seed kernels have been used as an antimalarial and anthelmintic.² As a part of our exploration of medicinal plant resources from Southeast Asia, we observed that the CH₂Cl₂ extract of the seed kernels of *C. crista* possessed significant antimalarial activity in mice infected with *Plasmodium berghei*.³ Further separation of the CH₂Cl₂ extract led to the isolation of seven new diterpenes, named caesalpinins C–G (**1–5**) and norcaesalpinins D and E (**6, 7**). In this paper, we report the isolation and structure elucidation of these new diterpenes together with antimalarial activity of the isolated compounds against *Plasmodium falciparum* FCR-3/A2 growth in vitro.

Results and Discussion

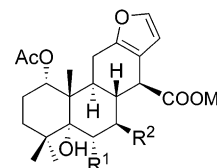
Air-dried seed kernels of *C. crista* Linn. were extracted with CH₂Cl₂ by overnight percolation at room temperature. In a test for antimalarial activity in vivo, the CH₂Cl₂ extract showed significant inhibition of parasitemia level



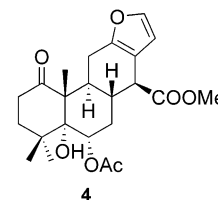
	R ¹	R ²	R ³	R ⁴
1	H	OAc	H	CH ₂
6	OAc	OAc	H	O
7	H	H	OH	O
8	OAc	H	H	O
9	H	OAc	H	O
15	OAc	OAc	H	CH ₂



	R ¹	R ²
2	H	OAc
5	OAc	H
12	H	H



	R ¹	R ²
3	OAc	H
17	H	OAc



(98.6%) at a dose of 10 mg/kg in mice infected with *P. berghei*. The CH₂Cl₂ extract was fractionated by silica gel column chromatography with a benzene/EtOAc gradient system to give nine fractions. Fractions 2–5 were further subjected to repeated silica gel column chromatography followed by normal- and reversed-phase preparative TLC to afford five new cassane-type diterpenes, caesalpinins C–G (**1–5**), and two additional norcassane-type diterpenes, norcaesalpinins D and E (**6, 7**), together with 11 known compounds, norcaesalpinins A–C (**8–10**), 2-acetoxy-3-deacetoxycaesaldekarin e (**11**),⁴ caesalmin B (**12**),⁵ caesaldekarin e (**13**),⁶ caesalpin F (**14**),⁷ 14(17)-dehydrocaes-

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Table 1. ¹H NMR (400 MHz) Data (δ) for Compounds **1–7** in CDCl₃ (J values in parentheses)

position	1	2	3	4	5	6	7
1	4.88 t (3.1)	4.90 br s	4.84 br s		5.26 d (2.9)	5.29 d (3.5)	4.94 t (2.9)
2	2.37 m, 2.15	2.03 m, 1.17 m	1.94 m, 1.75 m	1.91 m, 1.77 m	5.35 ddd (13.1, 4.9, 2.9)	5.54 t (3.5)	1.99 m, 1.77 m
3	4.95 t (3.1)	1.81 m 1.21 m	1.27 m 1.07 m	2.57 m 2.48 m	1.96 m 1.46 dd (13.1, 4.9)	5.18 d (3.5)	1.74 m 1.20 m
6	1.93 m 1.78 m	5.61 d (9.4)	5.36 dd (11, 5.5)	5.3 dd (11.5, 5.2)	2.38 dd (12.3, 5.0) 1.72 td (12.3, 2.7)	1.66 m 1.85 m	2.15 dd (11.1, 5.8) 1.67 td (11.1, 2.1)
7	1.21 m 2.06 m	4.77 dd (11.3, 9.4)	1.90 m 1.79 m	1.94 m 1.61 dd (23.2, 11.5)	4.67 td (11.0, 5.0)	2.20 m 1.80 m	4.38 td (11.1, 5.8)
8	2.36 m	2.09 m	2.17 m	2.13 m	2.05 m	2.38 td (12.3, 5.0)	2.39 dd (12.4, 11.1)
9	2.87 td (11.2, 5.6)	2.82 td (13.5, 8.8)	2.58 td (11.5, 6.0)	2.62 m	2.75 dt (13.3, 8.5)	3.20 td (12.3, 5.0)	3.08 td (12.4, 5.1)
11	2.54 dd (16.1, 11.2) 2.30 dd (16.1, 5.6)	2.53 m	2.42 ddd (16.6, 11.5, 2.7) 2.26 dd (16.6, 6.0)	2.38 m 3.35 dd (16.4, 4.9)	2.60 dd (17.9, 8.5) 2.49 dd (17.9, 8.5)	2.66 dd (17.0, 12.3) 2.52 dd (16.0, 5.0)	2.74 dd (17, 5.1) 2.53 dd (17, 12.0)
14		3.30 d (13.5)	3.30 d (9.5)	3.30 d (9.5)	3.23 d (13.3)		
15	6.45 d (2.5)	6.59 d (1.9)	6.14 d (2.0)	6.10 d (1.9)	6.60 d (1.7)	6.64 d (1.9)	6.65 d (1.9)
16	7.23 d (2.5)	7.30 d (1.9)	7.25 d (2.0)	7.20d (1.9)	7.30 d (1.7)	7.30 d (1.9)	7.33 d (1.9)
17	4.92 d (2.7) 5.12 d (2.7)						
18	1.11 s	1.17 s	1.13 s	1.10 s	1.15 s	1.13 s	1.07 s
19	1.15 s	1.18 s	1.17 s	1.20 s	1.21 s	1.25 s	1.11 s
20	1.16 s	1.22 s	1.19 s	1.40 s	1.26 s	1.32 s	1.24 s
1-OAc	2.04 s	2.16 s	2.11 s		2.18 s	1.99 s	2.10 s
2-OAc					1.99 s	2.11 s	
3-OAc	2.03 s					2.13 s	
6-OAc		2.16 s	2.09 s	2.09 s			
5-OH	3.27 br s	3.16 s	3.10 s		3.08 d (2.7)	3.30 d (3.0)	2.81 d (2.1)
7-OH							4.70 s
17-OCH ₃			3.75	3.74			

salpin F (**15**),⁷ 2-acetoxycasaldekalin e (**16**),⁷ 7-acetoxybonducellpin C (**17**),⁸ and caesalmin G (**18**)⁹ (Figure S1).

Caesalpinin C (**1**) was isolated as colorless amorphous solid with $[\alpha]_D^{25} +30.2^\circ$ (CHCl₃), whose molecular formula was determined to be C₂₄H₃₂O₆ by HRFABMS. IR absorptions at 3575 and 1735 cm⁻¹ indicated the presence of hydroxyl and carbonyl groups, respectively. The ¹H NMR spectrum (Table 1) displayed signals corresponding to three tertiary methyls, two oxygen-substituted methines, two aliphatic methines together with two protons of a 1,2-disubstituted furan ring (δ 7.23, 6.45), and two acetyl methyls. Moreover, the ¹³C NMR spectrum (Table 2) showed six olefinic carbons (δ 151.3, 142.2, 141.5, 119.1, 106.3, 104.3) and three oxygen-substituted carbons (δ 73.8, 76.9, 76.7) together with two ester carbonyl carbons. These ¹H and ¹³C NMR data were similar to those of 14(17)-dehydrocaesalpin F (**15**),⁷ except for the number of acetyl groups. Thus, **1** was considered to be a derivative of **15** having only one acetyl group instead of two in **15**. This was confirmed by analysis of the COSY, HMQC, and HMBC spectra. The locations of the acetyl groups were determined to be C-1 and C-3, on the basis of the long-range correlations of the ester carbonyl carbon at δ 169.4 (1-OCO) with the protons at δ 2.04 (1-OCOCH₃) and 4.88 (H-1) and of the ester carbonyl carbon at δ 169.3 (3-OCO) with the protons at δ 2.03 (3-OCOCH₃) and 4.95 (H-3). The relative configuration was assigned on the basis of NOEs between H₃-20 and H-1, H-2, H-6_{ax}, H-8, H-11, and H₃-19, and the coupling constants for H-1 ($J = 3.1$ Hz), H-3 ($J = 3.1$ Hz), and H-9 ($J_{9,11ax} = 11.2$ Hz). Thus caesalpinin C was **1**.

Caesalpinins D (**2**) and G (**5**) both were colorless amorphous solids with $[\alpha]_D^{25} +63.2^\circ$ (CHCl₃) and $[\alpha]_D^{25} +58.2^\circ$

Table 2. ¹³C NMR (100 MHz) Data (δ) for Compounds **1–7** in CDCl₃

position	1	2	3	4	5	6	7
1	73.8	74.9	75.8	212.6	73.8	73.9	75.2
2	26.5	22.4	21.8	38.1	66.5	66.2	22.7
3	76.9	32.6	32.5	35.3	35.2	77.4	29.9
4	41.6	39.4	38.5	38.5	40.9	43.3	38.4
5	76.7	83.7	78.2	80.7	81.6	76.7	77.9
6	23.4	72.6	72.0	72.7	30.1	26.5	33.6
7	22.3	82.3	32.5	32.2	80.2	20.5	67.2
8	35.1	44.4	34.0	34.0	29.8	44.0	51.0
9	38.7	32.6	36.6	37.9	33.4	40.2	38.6
10	43.8	47.6	45.2	56.1	45.9	45.6	43.6
11	26.5	21.6	21.8	23.4	21.4	23.2	22.8
12	151.3	151.3	150.0	151.5	151.3	165.8	166.6
13	119.1	113.9	113.9	112.5	113.8	120.2	119.8
14	142.2	41.5	48.0	48.1	41.1	195.5	198.4
15	106.3	107.8	108.7	108.5	107.8	106.8	106.3
16	141.5	142.0	141.4	141.0	141.8	143.3	143.4
17	104.3	173.2	173.7	173.9	174.4		
18	23.1	29.7	24.9	28.8	28.1	23.4	27.9
19	25.4	24.3	30.9	26.8	25.4	25.5	24.9
20	18.0	17.0	16.6	15.1	17.3	18.8	18.1
1-OCOCH ₃	21.4	21.3	21.4		20.9	21.2	21.5
1-OCOCH ₃	169.4	168.6	169		168.6	169.9	169.2
2-OCOCH ₃					21.0	21.4	
2-OCOCH ₃					170.3	170.1	
3-OCOCH ₃	21.1					20.9	
3-OCOCH ₃	169.3					169.6	
6-OCOCH ₃		21.3	22.2	21.7			
6-OCOCH ₃		169.7	170.1	169.2			
17-OCH ₃							

(CHCl₃), respectively. Their IR spectra indicated the presence of hydroxyl, γ -lactone, and ester groups, while the molecular formulas were identical, C₂₄H₃₀O₈, by HR-

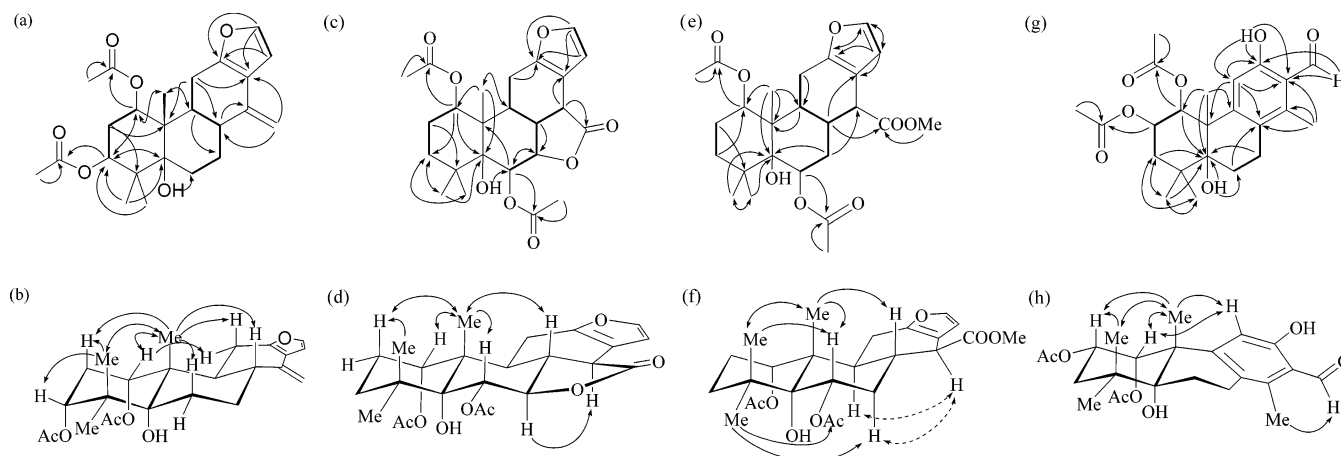


Figure 1. (a, c, e, g) Connectivities (bold lines) deduced by the COSY spectrum and key HMBC correlations (arrows) and (b, d, f, h) selected NOE (arrows) and ROESY (dashed arrows) correlations observed for **1**, **2**, **3**, and **8**.

FABMS. Their ^1H and ^{13}C NMR spectra were also similar and resembled those of caesalmin B (**12**),⁵ except for the presence of one more acetyl group. The locations of the additional acetoxy groups were C-6 and C-2, respectively, on the basis of the deshielding of H-6 (δ 5.61) and H-2 (δ 5.35), respectively, compared with those of **12** (H-6, δ 1.84; H-2, δ 1.92, 1.65). This was confirmed by HMBC correlations of the acetyl carbonyl carbons with the protons of acetyl methyl and of the protons of acetoxy-bearing carbons: the carbonyl carbon at δ 169.7 and protons at δ 1.99 and 5.29 in **2** and carbonyl carbon at δ 170.3 and protons at δ 1.99 and 5.35 in **5**. The relative configurations were determined on the basis of coupling constants and the results of difference NOE experiments. Thus, the structures of caesalpinins D and G were **2** and **5**, respectively.

Caesalpinin E (**3**) was isolated as a colorless amorphous solid, and its molecular formula was determined to be $\text{C}_{25}\text{H}_{34}\text{O}_8$ by HRFABMS. The ^1H and ^{13}C NMR spectra of **3** displayed signals corresponding to three tertiary methyls, two oxymethines, two aliphatic methines, a 1,2-disubstituted furan ring, two acetyl methyls, and a sharp singlet due to a carbomethoxy group. These data were similar to those of 7-acetoxybonducellpin C (**17**),⁸ suggesting that **3** was an isomer of **17** with respect to the location of the acetoxy group. Analysis of the COSY, HMQC, and HMBC spectra indicated that H-6 (δ 5.36) was deshielded and H-7 (δ 1.90, 1.79) shielded compared with H-6 at δ 2.17 and H-7 at δ 5.22 of **17**. Hence the acetoxy group of caesalpinin E (**3**) was at C-6, not at C-7, which was confirmed by the HMBC correlations (Figure 1e). The orientation at C-6 was determined to be α -OAc from the coupling constants of H-6 (dd, $J = 11, 5.5$ Hz), while that at C-14 was β -COOMe from the ROESY correlations of H-14 with H-7 and H-9 and the large J value (9.5 Hz) between H-14 and H-8. The relative configuration was confirmed by the results of difference NOE and ROESY experiments (Figure 1f). Thus, the structure of caesalpinin E was **3**.

Caesalpinin F (**4**) was isolated as a colorless amorphous solid whose molecular formula was $\text{C}_{23}\text{H}_{30}\text{O}_7$ by HRFABMS. The ^1H (Table 1) and ^{13}C NMR (Table 2) spectra were similar to those of caesalpinin E (**3**), except for the replacement of the oxymethine signal by that of a ketone carbonyl at δ_{C} 212.6, whose location was deduced to be on C-1 on the basis of the HMBC correlations of H₂-2, H₂-3, H-9, and H₃-20 with the ketone carbonyl carbon. The relative configuration was the same as that of **3** by difference NOE experiments. Thus, caesalpinin F (**4**) was the 1-keto derivative of **3**.

Table 3. Antimalarial Activity of the Isolated Compounds

compound	IC ₅₀ (μM)
1	0.76
2	0.80
3	6.50
4	0.65
6	2.0
7	0.09
8	0.80
9	0.26
10	5.0
11	0.098
12	0.80
13	4.0
15	0.20
16	6.50
17	0.60
18	> 10

Norcaesalpinin D (**6**) was isolated as a colorless amorphous solid and had an IR spectrum similar to those of norcaesalpinins A (**8**) and B (**9**). Its molecular formula $\text{C}_{25}\text{H}_{32}\text{O}_9$ was determined by HRFABMS. The ^1H (Table 3) and ^{13}C NMR (Table 2) spectra of **6** were also similar to those of **8** and **9**, but showed the presence of one more acetyl group (δ_{C} 77.4, δ_{H} 5.18) and one more oxymethine accompanied by the disappearance of signals due to one of three methylenes in **8** and **9**. The *O*-acetyl groups were attached to C-1, C-2, and C-3 by analysis of the COSY and HMQC spectra, which was further confirmed by HMBC analysis. NOE difference spectra showed that the relative configuration of **6** was the same as those of **8** and **9**. Thus, norcaesalpinin D (**6**) was 3-*O*-acetylnorcaesalpinin A.

The molecular formula of norcaesalpinin E (**7**) was $\text{C}_{21}\text{H}_{28}\text{O}_6$ by HRFABMS. The ^1H (Table 3) and ^{13}C NMR (Table 2) spectra were similar to those of bonducellpin C,⁸ except for the disappearance of signals due to one of two carbomethoxy groups, while the ^{13}C NMR spectrum indicated the presence of 19 carbons including a ketone carbonyl carbon. Thus, **7** was also a norditerpene. HMBC analysis indicated that the ketone carbon was at C-14; that is, **7** was a 17-norcassane-type diterpene. NOE difference spectra showed that the relative configuration of **7** was the same as that of norcaesalpinins A (**8**) and B (**9**). Thus, norcaesalpinin E (**7**) was 17-norbonducellpin C.

Of the new diterpenes, **6** and **7** are 17-norcassane-type diterpenes, which may be biosynthesized through oxidative decarboxylation of C-17 of cassane-type diterpenes such as 14(17)-dehydrocaesalpin F (**13**).⁷ Among the isolated compounds, 2-acetoxy-3-deacetoxycaesaldehydric acid (**11**),⁴ 14(17)-

dehydrocaesalpin F (**15**),⁷ 2-acetoxycaesaldehyd e (**16**),⁷ and 7-acetybonducellpin C (**17**)⁸ were obtained from natural sources for the first time.

The isolated compounds, except **5** and **14**,¹² were tested for their inhibitory activities against *P. falciparum* FCR-3/A2 growth in vitro.¹³ All of them displayed significant dose-dependent inhibition, norcaesalpinin E (**7**) exhibiting the most potent activity with an IC₅₀ value of 90 nM (Table 3). The IC₅₀ values of **7**, **9**, **11**, and **15** were less than that reported for a well-known antimalarial drug, chloroquine (IC₅₀, 283–291 nM).^{14,15}

Experimental Section

General Experimental Procedures. Optical rotations were recorded on a JASCO DIP-140 digital polarimeter. IR spectra were measured with a Shimadzu IR-408 spectrophotometer in CHCl₃ solutions. NMR spectra were taken on a JEOL JNM-LA400 spectrometer with tetramethylsilane (TMS) as an internal standard, and chemical shifts are expressed in δ values. HRFABMS measurements were carried out on a JEOL JMS-700T spectrometer, and glycerol was used as matrix. Column chromatography was performed with BW-802MH silica gel (Fuji Silysia, Aichi, Japan). Analytical and preparative TLC were carried out on precoated silica gel 60F₂₅₄ and RP-18F₂₅₄ plates (Merck, 0.25 or 0.50 mm thickness).

Plant Material. Seed kernels of *Caesalpinia crista* Linn. were collected at Polewali Mamasa, South Sulawesi Province, Indonesia, in September 2001 by one of the authors (F.A.). A voucher specimen (TMPW 21499) is preserved in the Museum for Materia Medica, Analytical Research Center for Ethnomedicines, Institute of Natural Medicine, Toyama Medical and Pharmaceutical University, Toyama, Japan.

Extraction and Isolation. A powder of air-dried seed kernels of *C. crista* (1.0 kg) was extracted with CH₂Cl₂ (3 L \times 2) at room temperature, overnight. The CH₂Cl₂ extract (151 g) was separated by silica gel column chromatography (8.5 \times 45.0 cm) with a benzene–EtOAc gradient system to give nine fractions. Fraction 1 (4.5 g) was fatty substances, as indicated by the NMR spectrum.

Fraction 2 (3.4 g) was rechromatographed (2.5 \times 40.0 cm) with a hexane–EtOAc gradient system to afford three subfractions (fraction 2-1, 55 mg; fraction 2-2, 109 mg; fraction 2-3, 117 mg). Subfraction 2-1 was subjected to preparative TLC with hexane–EtOAc (4:6) to give 2-acetoxy-3-deacetoxycaesaldehyd e⁴ (**11**, 8 mg). Subfractions 2-2 and 2-3 were also separated by preparative TLC with hexane–EtOAc (4:6) to give norcaesalpinin A (**8**, 32.9 mg) and norcaesalpinin C (**10**, 5.9 mg), respectively.

Fraction 3 (1.81 g) was rechromatographed (3.5 \times 17.7 cm) with a hexane–EtOAc gradient system to give four subfractions. Subfractions 3-1 and 3-2 were caesalmin B⁶ (**12**, 3.5 mg) and caesaldehyd e⁶ (**13**, 20.3 mg), respectively. Subfractions 3-3 (28.7 mg) and 3-4 (52.7 mg) were separately subjected to preparative TLC with 1% MeOH–CHCl₃; the former gave norcaesalpinin B (**9**, 19.7 mg), and the latter gave caesalpinin C (**1**, 19.7 mg) and caesalpinin F⁷ (**14**, 3.4 mg).

Fraction 4 (0.73 g) was rechromatographed (3.5 \times 14.8 cm) with a hexane–EtOAc gradient system to afford two subfractions (fraction 4-1, 235 mg; fraction 4-2, 490 mg). Subfraction 4-1 (235 mg) was subjected to preparative TLC with 0.5% acetone–CHCl₃ to give **13** (4.4 mg), 14(17)-dehydrocaesalpinin F⁷ (**15**, 7.7 mg), 2-acetoxycaesaldehyd e⁷ (**16**, 6.0 mg), 7-acetybonducellpin C⁸ (**17**, 4.4 mg), caesalpinin D (**2**, 0.8 mg), and norcaesalpinin D (**6**, 9.3 mg). Subfraction 4-2 (490 mg) was rechromatographed (1.5 \times 21.0 cm) over alumina with 1% acetone–CHCl₃ to afford two fractions (fraction 4-2-1, 53 mg; fraction 4-2-2, 35 mg). Each fraction was separated by preparative TLC with 0.5% acetone–CHCl₃, and **2** (5.9 mg), **6** (5.5 mg), **16** (4.9 mg), and **17** (10.2 mg) were obtained from fraction 4-2-1 and caesalpinin F (**4**, 5.4 mg), **9** (2.9 mg), norcaesalpinin E (**7**, 1.4 mg), and **17** (4.6 mg) were from fraction 4-2-2.

Fraction 5 (0.45 g) was rechromatographed (3.5 \times 15.8 cm) with a hexane–EtOAc gradient system to afford two subfractions. Subfraction 5-1 (241 mg) was separated by preparative TLC with 1% acetone–CHCl₃ to give **7** (7.6 mg) and three subfractions. Subfraction 5-1-1 was purified by reversed-phase preparative TLC with MeOH–CH₃CN–H₂O (2:1:1) to give caesalpinin G (**5**, 1.5 mg) and **16** (2.8 mg). Subfractions 5-1-2 and 5-1-3 were also purified by reversed-phase preparative TLC with MeOH–CH₃CN–H₂O (2:1:1) to give **2** (3.8 mg) and caesalmin G⁹ (**18**, 1.9 mg), respectively.

Caesalpinin C (1): colorless amorphous solid; [α]_D²⁵ +30.2° (c 0.11, CHCl₃); IR (CHCl₃) ν_{\max} 3575, 1735 cm⁻¹; HRFABMS *m/z* 417.2314 [calcd for C₂₄H₃₃O₆ (M + H)⁺, 417.2277]; ¹H and ¹³C NMR, see Tables 1 and 2.

Caesalpinin D (2): colorless amorphous solid; [α]_D²⁵ +63.2° (c 0.057, CHCl₃); IR (CHCl₃) ν_{\max} 3575, 1750, 1735 cm⁻¹; HRFABMS *m/z* 447.2025 [calcd for C₂₄H₃₁O₈ (M + H)⁺, 447.2031]; ¹H and ¹³C NMR, see Tables 1 and 2.

Caesalpinin E (3): colorless amorphous solid; [α]_D²⁵ +126.0° (c 0.02, CHCl₃); IR (CHCl₃) ν_{\max} 3575, 1735 cm⁻¹; HRFABMS *m/z* 463.2317 [calcd for C₂₅H₃₅O₈ (M + H)⁺, 463.2332]; ¹H and ¹³C NMR, see Tables 1 and 2.

Caesalpinin F (4): colorless amorphous solid; [α]_D²⁵ +47.0° (c 0.081, CHCl₃); IR (CHCl₃) ν_{\max} 3575, 1730, 1710 cm⁻¹; HRFABMS *m/z* 419.2051 [calcd for C₂₃H₃₁O₇ (M + H)⁺, 419.2027]; ¹H and ¹³C NMR, see Tables 1 and 2.

Caesalpinin G (5): colorless amorphous solid; [α]_D²⁵ +58.2° (c 0.063, CHCl₃); IR (CHCl₃) ν_{\max} 3575, 1750, 1735 cm⁻¹; HRFABMS *m/z* 447.2009 [calcd for C₂₄H₃₁O₈ (M + H)⁺, 447.2019]; ¹H and ¹³C NMR, see Tables 1 and 2.

Norcaesalpinin D (6): colorless amorphous solid; [α]_D²⁵ +3.3° (c 0.09, CHCl₃); IR (CHCl₃) ν_{\max} 3575, 1740, 1715 cm⁻¹; HRFABMS *m/z* 477.2106 [calcd for C₂₅H₃₃O₉ (M + H)⁺, 477.2125]; ¹H and ¹³C NMR, see Tables 1 and 2.

Norcaesalpinin E (7): colorless amorphous solid; [α]_D²⁵ +84.7° (c 0.01, CHCl₃); IR (CHCl₃) ν_{\max} 3575, 1735, 1715 cm⁻¹; HRFABMS *m/z* 377.1946 [calcd for C₂₁H₂₉O₆ (M + H)⁺, 377.1964]; ¹H and ¹³C NMR, see Tables 1 and 2.

Antimalarial Activity. To determine the antimalarial activity of each isolated compound, a malaria parasite, *P. falciparum* FCR-3/A2 clone, was propagated in a 24-well culture plate in vitro in the presence of a wide concentration range of each compound, following the procedure described previously (Budimulja et al., 1997). Each compound was separately dissolved in DMSO to obtain a 10⁻² M stock and kept at –20 °C until used. The parasite growth was monitored by making a blood smear every day. The concentration response parasite growth data were analyzed by a linear regression function using the Sigma-plot 2000 computer program to determine the 50% inhibitory concentration (IC₅₀). The IC₅₀ value is defined as that concentration of compound producing 50% growth inhibition relative to untreated control.

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Supporting Information Available: Figures S1 and S2 showing the structures of the known compounds isolated from *C. crista* and possible biogenesis of norcaesalpinins C and D are available free of charge via the Internet at <http://pubs.acs.org>.

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